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NUCLEAR PACKING EFFICIENCY

by

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NUCLEAR PACKING EFFICIENCY

BACKGROUND OF THE INVENTION

FIELD OF THE INVENTION

5 The present invention relates to cell biology, and more particularly to characterizing a cell by its nuclear packing efficiency (NPE).

BACKGROUND INFORMATION

10 Virtually all eucaryotic cells have a nucleus, a compartment enclosed by the nuclear membrane and containing most of the cell's DNA, as well as other nuclear components. Aberrations in the size and shape of the nucleus have long been recognized as an indication of cancer and other diseases, although their
15 characterization was previously limited to microscopic observation.

 Since then, attempts have been made to characterize cells by quantitating and comparing various nuclear components. For example, measurements of DNA,
20 RNA and nuclear protein have been compared with each other. One study measured the quantity of DNA and tried to correlate it with measurements of the size of the nucleus, as measured by light scatter, time of flight and area. However, these correlations have been hampered by
25 unreliable indirect estimates of the nuclear volume.

 Indirect estimates of volume from diameter measurements using light scatter have worked best assuming uniform spherical particles of a certain size range and having a relatively high index of refraction.

But this technique becomes less accurate when applied to nonideal biological samples outside the optimal range of measurement. Other measurements, termed "time-of-flight" or TOF, measure the size of particles as a flowstream carries the particles across a beam of light. However, this technique is subject to many limitations, including sensitivity to fluctuations in the speed of the particles and variations from the relative orientation of the particles, and only yields a measurement of one axis of the three-dimensional particle.

Still other indirect measurements estimate nuclear volume based on the cross-sectional areas of the nucleus. But these measurements, in turn, can be limited by variability in the staining and mounting techniques used on the nuclei. In particular, confocal microscopes have been used to measure the area of stained DNA in the nucleus, summing up successive cross-sections to obtain a measure of the total DNA. By assuming that the nuclear volume is proportional to the stained DNA, this technique then yields an estimate of the nuclear volume. However, this technique fails to account for the granularity of DNA within in the nucleus, and ignores the varying contribution to nuclear volume from the other components of the nucleus: RNA, nuclear proteins, nuclear lipids, nuclear envelope and nuclear water.

Thus, previous techniques fail to meet the need for satisfactory measurements of nuclear volume in combination with a useful correlation with other measurements to characterize the condition of nuclei and the cells as a whole. The present invention satisfies this need and provides related advantages as well.

SUMMARY OF THE INVENTION

The nucleus of a cell is a highly organized structure, allowing precursor materials to pass through pores in the nuclear envelope into the nucleus, and
5 nuclear products to be transported out to the cytoplasm. The complex nuclear machinery--nucleic acids, proteins, lipids and other components--are tightly packaged within the volume of the nucleus.

As with most organized structures in nature,
10 the nucleus assumes a shape for compact packaging of its components for optimal efficiency. When a cell becomes diseased, such as in malignant cells, the nuclear organization breaks down. For example, the DNA loses its ability to fold efficiently around histone proteins into
15 organized structures called nucleosomes. The protein content also changes, as well as other biochemical components of the nucleus. The volume of the nucleus becomes forced to increase to accommodate this disorganization. Thus, the efficiency of this packing is
20 a characteristic of the nucleus--and a useful indication of the condition of the cell as a whole.

The present invention provides methods and devices for determining the nuclear packing efficiency (NPE) of a cell by measuring the spatial displacement of
25 the nucleus (SDN), for example by using flow cytometry to measure electronic nuclear volume (ENV). When the method is applied to procaryotes or viruses, the SDN can be considered the volume of the surrounding particle, which is the procaryotic cell or the virus itself. One or more
30 biochemical components (BCs) of the nucleus are also measured, such as nucleic acids, nuclear protein, nuclear

lipids or nuclear water. An NPE is then determined by correlating the values measured for BC and SDN.

A variety of techniques can be used to correlate the BC and SDN to yield an NPE. Polynomial fitting can be used, from the ratio BC/SDN to more complex expressions such as $NPE = k_1(BC)^a / (SDN)^b + k_2(BC)^c + k_3(SDN)^d + k_4$. Graphical methods are particularly useful for evaluating NPEs for a population of cells and for identifying distinct subpopulations of cells. Subpopulations can then be characterized in terms of their geometric parameters, such as diameter, eccentricity and gradient line slope.

Once determined, NPEs are useful for identifying cells having a phenotype of interest. For example, cells can be identified by tissue source and by the sex and species of the organism, as well as by various states of differentiation and stages in the cell division cycle and apoptosis. NPEs can also identify cells having various disease states as they differ from their normal states, particularly neoplastic cells exhibiting aneuploidy, distinguishing among benign, malignant and metastatic cells, thus enabling the diagnosis and prognosis of cancer.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1a to 1c show NPE contours (electronic nuclear volume v. DNA fluorescence) for normal human cells from surface oral epithelium (Fig. 1a), intestine (Fig. 1b) and thyroid (Fig. 1c). NPE contours for other normal human cells are shown in Figures 2a, 3a and 5a.

Figures 2a to 2e compare NPE contours for human lymph node and breast cells in various states: normal human lymph node cells (Fig. 2a and perspective view Fig. 2b), cells from a benign breast tumor (Fig. 2c), cells taken from a malignant primary tumor from breast tissue (Fig. 2d) and cells taken from metastasizing breast cells from the tumor shown in Figure 2d to the lymph node (Fig. 2e).

Figures 3a to 3c compare NPE contours for cells from human colon in various states: normal colon cells (Fig. 3a), cells from a primary colon tumor (Fig. 3b) and metastatic cells taken from an end point of a surgical resection performed to remove the tumor shown in Figure 3b (Fig. 3c).

Figures 4a to 4f show NPE contours for human cells taken from four cancerous tissue sources: gastric (Fig. 4a and perspective view Fig. 4b), prostate (Fig. 4c), ovarian (Fig. 4d and perspective view Fig. 4e) and lung (Fig. 4f).

Figures 5a to 5c illustrate NPE contours for human lymphocytes in various states: normal lymphocytes (Fig. 5a), leukemic lymphocytes (Fig. 5b), and activated lymphocytes (Fig. 5c)

Figure 6a shows an NPE contour for cells from mouse cell line P388. Figure 6b shows NPE contours for normal and apoptotic cells from a WEHI-231 murine B lymphoma cell line (Fig. 6b and perspective view 6c).

Figure 7 shows a block diagram for a device for performing the method of the invention.

In the figures, the following reference numbers are used:

- | | | |
|----|----|---|
| 5 | 1 | trout red blood cell nuclei (TRBC)
internal standard |
| | 2 | diploid G_0/G_1 cluster |
| | 2a | cluster of diploid G_0 cells |
| | 2b | cluster of diploid G_1 cells |
| 10 | 2c | cluster of aneuploid G_0 cells |
| | 2d | second cluster of aneuploid G_0 cells |
| | 2e | cluster of activated lymphocytes |
| | 2f | cluster of apoptotic cells |
| | 3 | diploid cells in S phase |
| 15 | 3c | cluster of aneuploid S cells |
| | 4 | diploid G_2 +M cluster |
| | 4a | cluster of diploid G_2 cells |
| | 4b | cluster of diploid M cells |
| | 4c | cluster of aneuploid G_2 +M cells |
| 20 | 5 | normal diploid NPE line |
| | 5c | aneuploid NPE line |
| | 5d | second aneuploid NPE line |
| | 5f | apoptotic NPE line |
| 25 | 6 | gradient line for cluster |
| | 6e | gradient line for activated
lymphocytes |

Not all clusters may be visible in a given contour due to the particular threshold values selected for display and printout.

DETAILED DESCRIPTION OF THE INVENTION

5 The components of the nucleus of a cell are packaged within the volume of the nucleus. The efficiency of this packing is a characteristic of the nucleus--and a useful indication of the condition of the cell as a whole. Thus, the present invention provides a
10 method for determining the nuclear packing efficiency (NPE) of a cell. The method measures the spatial displacement of the nucleus (SDN) and one or more biochemical components (BCs) of the nucleus. The NPE is then determined by correlating the values of SDN and the
15 BCs.

"Spatial displacement of a nucleus" (SDN) as used herein means the volume of space that is occupied by the nucleus. By occupying that volume, the nucleus can be said to displace any non-nuclear matter that would
20 have otherwise occupied that space.

"Nucleus" as used herein generally means the organelle surrounded by the cytoplasm of the eucaryotic cell that contains the chromosomal DNA. The term encompasses the inner and outer walls of the nuclear
25 envelope and their associated proteins.

In addition to eucaryotic cells, the NPE can also be applied to procaryotic cells, which lack a nucleus in the eucaryotic sense. In this context the term "nucleus" is therefore used to refer to the particle
30 size of the entire procaryotic cell itself. Thus, •

references to "nuclear" volume or spatial displacement can refer to the volume or spatial displacement of the procaryotic cell. Similarly, biochemical components of the "nucleus" can also refer to cellular components of
5 procaryotic cells.

Furthermore, the NPE can be applied to viruses. Because viruses are not considered cells, the term "nucleus" in this context can refer to the particle size of the entire virus or of a portion such as a capsid.
10 Thus, references to spatial displacement volume of a "nucleus" and biochemical components of the "nucleus" in the context of viruses should be understood to refer to the volume and components of the viral particle as a whole. Volume determination for viruses is well known in
15 the art (DeBlois and Wesley, J. Virol. 23:227-233 (1977)).

The electronic cell volume (ECV) method is a particularly useful method for measuring SDN, yielding an "electronic nuclear volume" (ENV). As used herein, the term "ENV" means the volume of the nucleus as determined
20 by an ECV method. When determining ECV, particles such as nuclei are suspended in a conducting fluid, which is passed through a small aperture. An electric field, either DC or AC, is then applied across the aperture, causing current to flow through the aperture. When a
25 particle passes through the aperture, the current is disrupted, causing a measurable pulse in the current. This pulse can be used to count individual particles as they pass through the aperture. The dimensions of the pulse can also be related to the size of the particle.

30 Refinements of the basic ECV method include shaping the inlet and outlet volumes of the measuring aperture to reduce edge effects and to produce a linear

relationship between the measured change in current and the volume of the particle. A particularly useful method for measuring ENV is described in U.S. Patent No.

4,818,103, to Thomas, which describes an improved flow
5 cytometry aperture.

Compared to previous methods, such as forward angle light scatter, the ECV method is not significantly affected by the shape of the particles as they are carried in the fluid. This is particularly important
10 when the particles can be nonspherical, such as biological particles injected into the center of the stream. A useful method uses a time-of-flight (TOF) signal to account for such changes, as described in U.S. Patent No. 4,298,836 to Groves. The TOF measurements can
15 be taken from the optical fluorescent pulse generated by the nucleus as it traverses an excitation light beam, or directly from the ECV pulse. The width of the pulse is related to the long axis of the particle.

An SDN can be expressed in terms of volume such
20 as cubic microns or cubic millimeters. An SDN can also be expressed in terms of a signal generated from an ECV device, whether analog or digital. In the Figures and Examples below, the ENV may be expressed as an 8-bit value, providing 256 channels of resolution.

25 Nevertheless, the conversion from channels to cubic microns is provided in each figure. However expressed, the SDN is then correlated with BC measurements to determine the NPE. It should also be noted that the method does not require that the SDN and the BCs be
30 measured in any particular order.

"Biochemical component" (BC) as used herein means any identifiable substance in the cell that can be quantitated in physical terms. For example, a BC can be measured in terms of amount, volume or area, such as surface or cross-sectional area. Examples of BCs include various nucleic acids such as DNA and RNA, proteins, lipids and nuclear water, as well as mixtures and subsets of individual BCs.

DNA in a nucleus can be conveniently measured by flow cytometry methods. Flow cytometry methods are generally described in M.G. Ormerod, Flow Cytometry (BIOS Sci. Pubs., 2nd ed. 1999, and referenced cited therein). When nuclei are treated with a DNA stain, such as a fluorescent stain, the quantity of DNA in the nuclei can be measured by detecting the amount of staining or fluorescence. DNA stains include propidium iodide, acridine orange, ethidium bromide, quinacrine, mithramycin, chromomycin A3 and 4',6-diamidino-2-phenylindole (DAPI) (see Krishan, J. Cell Biol. 66:188-193 (1975); Darzynkiewicz et al., Cytometry 5:355-363 (1984); Kapuscinski, Biotechnic & Histochem. 70(5):220-233 (1995)) (see Example I). Automated microscopes that measure absorption or fluorescence can also be used with DNA staining to measure the quantity of DNA in the cell nucleus (Tanke et al., J. Histochem. Cytochem. 27:84-86 (1979); Bjelkenkrantz, Histochemistry 79(2):177-91 (1983)).

Indirect measurements of DNA are possible by measuring the proteins and other components associated with DNA structures in the nucleus. Chromosomal DNA is coiled around octomers of histone proteins (two each of H2A, H2B, H3 and H4) to form a complex termed a nucleosome. Together with nonhistone chromosomal

proteins, DNA-histone structures form fibers collectively termed chromatin, which is itself divided into metabolically active euchromatin and transcriptionally inert heterochromatin. Measurement of any of these DNA-associated proteins can therefore be a useful measure of DNA content. For example, labeled antibodies that specifically bind to any of these proteins are particularly useful for such indirect measurements of nuclear DNA. In particular, histones be measured by flow cytometry and automated microscopes using antibodies against specific histones (Miller et al., Hybridoma 12(6):689-698 (1993)). Histones can also be quantitated by electrophoretic examination using a silver stain (Tsutsui et al., Anal. Biochem. 146(1):111-117 (1985)).

RNA has similarly been measured with flow cytometry using acridine orange staining (Piwnicka et al., Cytometry 3(4):269-75 (1983)). RNA can also be measured by cytophotometric image analysis using methyl green-pyronin Y stain (Schulte et al., Histomchem. J. 24(6):305-310 (1992)). As with DNA, RNA-related proteins, such as nuclear proteins related to RNA transcription, can be used to measure RNA indirectly.

The nucleolus is another nucleic-acid-related BC, a large structure where large numbers of rRNA copies are transcribed and immediately packaged with ribosomal proteins to form ribosomes. Thus, antibodies against any of the nucleolus machinery can provide a useful BC for measurement.

Lipids that are BCs include any measurable lipids in the nucleus such as nuclear envelope lipid, whether in the inside or outside walls. These lipids can be measured directly or indirectly by measuring nuclear-

envelope-associated proteins. Outer wall nuclear membrane proteins include ribosomes. Proteins associated with the inner wall of the nuclear membrane include nuclear pore proteins, nuclear lamina proteins and lamina-associated polypeptides. Fluorescent lipophilic dyes (Collas et al., Dev. Biol. 169(1):123-35 (1995)) and monoclonal antibodies that recognize the nuclear pore antigens (Matsouoka et al., Biochem. Biophys. Res. Commun. 254(2):417-423 (1999)) can be used to measure various components of the nuclear envelope and to provide a measure of lipid content, as well.

Although proteins have been discussed as a means to indirectly measure nucleic acids and lipids, nuclear proteins can be BCs in their own right. Nuclear proteins can be detected by nonspecific staining using FITC fluorescence in combination with flow cytometry (Roti et al., Cytometry 3(2):91-96 (1982)). Another method is to measure dinitrofluorobenzene (DNF) absorbance using cytophotometric image analysis (Cohn et al., Histochemistry 79(3):353-364 (1983)). Yet another method is to measure protein-bound sulfhydryl groups with the fluorescence of AEDANS (Schabronath et al., Cytometry 11(3):333-340 (1990)).

Nuclear Matrix Proteins (NMPs) and other specific nuclear proteins can also be measured as BCs. NMPs can be measured using antibodies prepared against purified protein molecules located in the nuclear matrix. As with the antibodies discussed above, they can then be conjugated with fluorescent stains and used with flow cytometry or immuno-histochemical staining to detect specific NMPs (Hughes et al., Am. J. Clin. Path. 111(2):267-274 (1999)).

Nuclear water and associated non-organic salts make up the remainder of the volume in the nucleus. Nuclear water can be measured by ENV when performed at isotonic and iso-osmotic conditions.

5 As disclosed above, an NPE is determined by correlating the SDN and BC values. This correlation may be achieved by a variety of mathematical functions and operations. For example, the NPE can be determined by using a general polynomial function such as

$$10 \quad NPE = k_1(BC)^a / (SDN)^b + k_2(BC)^c + k_3(SDN)^d + k_4$$

In this formula, k_1 , k_2 , k_3 , k_4 , a , b , c and d are individually preselected constants and k_1 is not zero. Particularly useful values for k_1 , k_2 , k_3 , k_4 , a , b , c and d are 2, 1, 1/2, 0, -1/2, -1, and -2 independently, with
 15 the proviso that k_1 is not zero. A specific application of the general formula is where each of k_1 , a and b are 1 and each of k_2 , k_3 and k_4 are zero, resulting in the ratio

$$NPE = BC / SDN$$

Determination of an NPE is not limited to measuring and
 20 correlating a single BC, however.

A second biochemical component (BC_2) can be measured as a further step in the method. The BC_2 can be any of the BCs disclosed above, such as nucleic acids, lipids, proteins and nuclear water, so long as it is
 25 different from the first BC. BC_2 can then be incorporated into the determination of NPE by a variety of formulas using the expression $k_5(BC_2)^e$, where k_5 and e are each preselected constants such as 2, 1, 1/2, 0, -1/2, -1 and -2. For example, $k_5(BC_2)^e$ can be added to the value of BC

in the general formula, so that the general formula becomes

$$\text{NPE} = k_1(\text{BC} + k_5(\text{BC}_2)^e)^a / (\text{SDN})^b + k_2(\text{BC} + k_5(\text{BC}_2)^e)^c + k_3(\text{SDN})^d + k_4.$$

- 5 Similarly, $k_5(\text{BC}_2)^e$ can be added to the value of SDN in the general formula to obtain

$$\text{NPE} = k_1(\text{BC})^a / (\text{SDN} + k_5(\text{BC}_2)^e)^b + k_2(\text{BC})^c + k_3(\text{SDN} + k_5(\text{BC}_2)^e)^d + k_4.$$

- Furthermore, the NPE obtained by the general polynomial
10 formula can be multiplied by $k_5(\text{BC}_2)^e$:

$$\text{NPE} = k_5(\text{BC}_2)^e \times (k_1(\text{BC})^a / (\text{SDN})^b + k_2(\text{BC})^c + k_3(\text{SDN})^d + k_4).$$

The polynomial expressions above are merely intended to illustrate the range of useful correlations possible between BCs and SDN. Specific examples of NPE

- 15 correlations include the following:

$$\text{NPE} = \text{DNA}/\text{ENV}$$

$$\text{NPE} = (\text{fluorescence of DNA})/\text{ENV}$$

$$\text{NPE} = \text{DNA}/\text{ENV}^{1/2}$$

$$\text{NPE} = \text{DNA}/(\text{procaryotic cell volume})$$

20 $\text{NPE} = \text{RNA}/(\text{volume of viral particle})$

$$\text{NPE} = \text{NMP}/\text{ENV}$$

$$\text{NPE} = (\text{mass of nucleosome})/\text{ENV}$$

$$\text{NPE} = (\text{volume of nuclear envelope})/\text{ENV}$$

$$\text{NPE} = (\text{DNA} - \text{RNA})/\text{ENV}$$

25 $\text{NPE} = (\text{DNA} + \text{RNA})/\text{ENV}^2$

$$\text{NPE} = (\text{DNA} \times \text{RNA})/\text{ENV}^2$$

$$\text{NPE} = \text{DNA}/(\text{ENV} - (\text{nuclear water}))$$

$$\text{NPE} = (\text{DNA} + \text{nuclear matrix protein})/\text{ENV}$$

$$\text{NPE} = \text{DNA}^{-1}/(\text{ENV} + \text{RNA})^{-1}$$

$$\text{NPE} = (\text{DNA} + \text{RNA})/\text{ENV}^2$$

$$\text{NPE} = 2(\text{DNA})^2/\text{ENV} + 0.5(\text{DNA}) - \text{ENV}^2 + 5$$

$$5 \quad \text{NPE} = (\text{DNA} + \text{RNA})^2/\text{ENV} - (\text{DNA} + \text{RNA})^{-1/2} + \text{ENV}$$

Mathematical variations of these correlations between BC and SDN can include trigonometric, logarithmic and exponential and other transcendental functions while not falling within the strict formula of the polynomial
 10 itself. These variations should be considered equivalent so long as they mathematically relate BC and SDN in a substantially similar way to achieve a substantially similar correlation that is a useful measure of NPE. Thus, determining NPE is not limited to functions,
 15 however, but can be determined using a number of more sophisticated methods.

Graphical methods provide a powerful tool for determining and visualizing NPEs (see Examples II.D and III). As an example, a BC and SDN can be measured for a
 20 cell and plotted on separate axes for BC and SDN, resulting in a datapoint for the cell. The term "datapoint" means herein any mathematical correlation between two or more values. Thus, a datapoint for a cell can also correlate DNA, RNA and SDN. The term can refer
 25 to a graphed or plotted point, but can also refer to any representation of the values of BC and SDN, whether displayed visually, represented numerically or stored in a computer's memory. However, each datapoint's values should represent a single cell or small group of cells.

Datapoints for multiple cells can also be plotted, either separately or on the same plot. If on the same plot, the datapoints can be represented by a variety of graphical methods. Scattergrams simply show each datapoint on the same graph, where the density of datapoints reflects the number of cells having BCs and SDNs of a certain value or range. Contour plots can be used illustrate the frequency of datapoints in a certain range by plotting a contour or surface, where a separate axis is used to indicate frequency.

An example of a graphical representation of datapoints is Figure 2b, which shows data from normal human lymph node. The horizontal axis is DNA fluorescence, reflecting the amount of DNA in each nucleus, and the depth axis is ENV, reflecting nuclear volume. The vertical axis is the number of cells having a datapoint at each given value on the horizontal and depth axes. Thus, a frequency contour is provided for ENV v. DNA.

As shown in this perspective view, the majority of the lymph node cells are in a single peak representing a cluster of cells 2 that are diploid G_0/G_1 cells. The second highest peak represents a cluster of trout red blood cell nuclei (TRBC) 1 added to the sample as an internal control (see Example I.C.). A much smaller peak is also visible representing a cluster of diploid G_2+M cells 4 in the sample of lymph node cells.

The same data can be represented in a contour graph. In Figure 2a, the horizontal axis is DNA fluorescence and the vertical axis is ENV. The frequency of cells having each ENV v. DNA data point is shown by

contour lines, representing lines of equal frequency, much as a topographic map uses lines of equal elevation. As before, the cluster of TRBC internal standard 1 is seen, as well as clusters representing the two
 5 subpopulations of lymph node cells: diploid G_0/G_1 cells 2 and diploid G_2+M cells 4.

Surprisingly, a line 5 can be drawn from the origin through the centers of the clusters 2 and 4. This is because the two clusters share a common correlation
 10 between ENV and DNA, even though the individual values for ENV and DNA differ by about two-fold. In short, the two clusters of lymph node cells share a common NPE that is characteristic for lymph node cells. In graphical terms, where a line has the formula $BC = NPE(SDN)$, the
 15 slope of the line (NPE) is BC/SDN . Alternatively, the line can have the formula $SDN = NPE(BC)$, where the slope (NPE) is SDN/BC . A line or substantially linear curve representing NPE can be determined graphically by hand, by calculator or determined by a variety of mathematical
 20 and statistical software packages. Thus, an NPE can be determined graphically for datapoints representing multiple cells.

An added advantage to representing the datapoints graphically is that distinct clusters of
 25 datapoints can be identified, each representing subpopulations of cells in the original sample. For example in Figure 2a, clusters 2 and 4 are readily identified with separate subpopulations of cells. The term "cluster" means herein any subset of the entire set
 30 of datapoints plotted for a population of cells or for a representative number of cells in a preselected population. Similarly, the term "subpopulation" means

herein the cells represented by the datapoints in a cluster. Thus, in Figure 6a, separate subpopulations of cells can be discerned by clusters 2a, 2b, 3, 4a and 4b. It should be noted, however, that when virtually all the datapoints form a well-defined group, cluster 2 in Figure 1a for example, the terms cluster and subpopulation may still apply.

More formally, a cluster can be defined as a neighborhood of datapoints that are adjacent to a local maximum of datapoints and characterized by decreasing frequency as the distance from the local maximum increases. If desired, a frequency threshold or cut-off can be used to further resolve the separation between clusters, and the subpopulations represented by the clusters.

Geometric parameters can then be used to describe the characteristics of each cluster. The term "geometric parameter" means herein any geometric or mathematical property of a cluster. The center of a roughly circular cluster is a parameter that can be defined as the average center point, the centroid or the local maximum. The slope of a line passing through the origin and the center of a cluster is a particularly useful parameter. Where the shape of the subpopulation contour is ellipsoid, other useful geometric parameters include the eccentricity, the maximum range of the major axis, the maximum range of the minor axis and the standard deviations of the major and minor axes. Another useful geometric parameter is the perimeter of the cluster when represented graphically at a predetermined threshold value.

Other mathematical variations of these geometrical parameters for clusters can include a variety of polynomial, trigonometric, logarithmic, exponential and other transcendental functions, while not falling within the list of parameters described above. These variations should be considered equivalent so long as they mathematically describe a geometrical feature of the cluster in a substantially similar way to achieve a substantially similar description that is a useful description of the cluster.

A gradient line is another particularly useful geometric parameter, indicating the general tilt of an ellipsoid or elongated cluster. The term "gradient line" means the line passing orthogonally through the direction of highest slope in a cluster. Examples of gradient lines are the lines marked 6 in Figure 2d. The line orthogonal to the gradient line can also be a useful geometric parameter of a cluster. Although a gradient line can often be hand-drawn by inspection, the line can be determined more precisely by performing a linear regression of the datapoints in the subpopulation.

Having generated a contour of datapoints and identified clusters representing subpopulations of cells in the sample, the geometric parameters identified can then be used to identify different cells within a population of cells by identifying the cell if the cell's NPE is within at least one predetermined NPE range. Separately or concurrently, the method can further involve segregating the identified cell from non-identified cells. The term "segregating" herein means to separate the cells into distinctly separate areas or containers and are not in their original state or in a uniform mixture.

It should be emphasized that the methods for determining an NPE for a population of cells are not limited to graphical, but can be performed equally well numerically. Thus, each of the steps of the disclosed method for determining an NPE for a population of cells--including determining a datapoint for BC and SDN, identifying a cluster of datapoints and determining an NPE according to a preselected geometric parameter--can be performed numerically. For example, these steps can be performed by a computer without necessarily representing the data in graphical form. Thus, disclosed applications for NPEs and NPE contours should be understood to apply equally to both.

NPEs and NPE contours can be used to identify cells having a phenotype of interest. Cells can be identified by comparing their NPEs with other NPEs. For example, an NPE of a cell of interest can be compared with a predefined range of NPEs for a reference population of cells, or against other cells in the sample. As a result, NPEs and NPE contours are a useful characteristic to identify the phenotype of cells in a sample.

A particular use for NPEs is to determine the sex of an organism by its cells. In the somatic cells of many species, cells from female animals have two X chromosomes, while cells from male animals have one X and a smaller Y chromosome. As a result, male cells have less DNA content than female cells, but with comparable nuclear volumes. This difference is reflected by a lower packing efficiency. In Figure 2a, cluster 2 tends to have a lower DNA content (DNA fluorescence channel closer to 76) when cells are from males and a higher DNA content (DNA fluorescence channel closer to 80.5) when the cells

are from females. These changes are reflected in a higher slope (decreased NPE) for males and lower slope (increased NPE) for females. Thus, cells from animals of different sexes can be distinguished. The method is
5 equally applicable in other animals, such as certain waterfowl, where the sex chromosomes are reversed.

NPEs can also be used to determine whether cells are from different tissues. For example, Figures 1a, 1b, 1c, 2a, 3a and 5a show NPE contours for cells
10 from different tissues, each having characteristic NPEs. Thus, cells of unknown origin can also be identified by comparing their NPEs with NPEs of known tissues. Similarly, cells in different stages of differentiation can be identified by their NPEs (see Example III.C.).

15 A further use for NPEs is to determine whether cells are from different species. In Figure 1a, for example, the NPE for nuclei from trout are distinguished from an NPE for human lymph cells. Similarly, the NPEs of the clusters in Figure 6, representing cells from a
20 mouse cell line, are distinguishable from the NPE in Figure 1a. Thus, cells from different species can be distinguished by their NPEs.

Cells in different stages of the cell division cycle can also be identified using NPE methods. Normally
25 dividing cells undergo a well-defined series of stages to coordinate cell division into two daughter cells and the corresponding replication of DNA necessary to maintain a complete set of chromosomes for the daughter cells. During S ("synthesis") phase, a cell replicates its
30 nuclear DNA, doubling the DNA content of the nucleus. After resting during G₂ phase, the nucleus divides during

M ("mitosis") phase, evenly separating the replicated DNA, followed by cytokinesis, where the cell itself divides into separate daughter cells. Cell division is then followed by the G_0 and G_1 phases before initiating S phase again. As shown in Example **III.D**, NPEs can be used to identify cells in different stages of the cell cycle.

Because the NPE of a cell is maintained during the cell cycle so rigorously, it follows that disruptions of the condition and normal growth of a cell will be reflected in its NPE. Thus, the NPE can be used to identify cells in an apoptotic state, when the cells undergo apoptosis, programmed cell death (see Example **III.E**).

Furthermore, NPEs can be used to identify an abnormal conditions such as a pathology or disease state. Disease states include genetic diseases such as sickle cell anemia. For example, the extra chromosomes present from Down's syndrome and Klinefelter's syndrome can be detected using NPEs. Similarly, genetic anomalies associated with autoimmune diseases can also be detected.

NPEs are particularly useful for identifying a neoplastic state. As used herein, the term "neoplastic" means characterized by formation and growth of abnormal tissue that grows more rapidly than normal. Neoplastic tissue can show partial or complete lack of structural organization and functional coordination with normal tissue. Neoplastic cells are often characterized by aneuploidy. The term "aneuploidy" herein means having an abnormal number of chromosomes. Aneuploid cells are contrasted with cells having a normal number of chromosomes, although the precise number can vary depending on the stage of division in the cell division

cycle. Thus, the method can be useful for distinguishing neoplastic cells from normal tissue.

As discussed in Example **III**, normal cells (see Figure **2a**) can be characterized by relatively circular clusters, while neoplastic cells can be characterized by relatively elongated clusters (see Figures **2c**, **2d**, and **2e**). The clusters can become elongated due to the presence of aneuploid cells, labeled **2c**, **2d** and **4c**. In turn, these clusters of aneuploid cells can give rise to one or more aneuploid NPE lines **5c**, **5d** (see Figures **2d**, **2e**).

Among neoplastic cells, benign tumors can have clusters with relatively vertical gradient lines **6** (see Figure **2c**), while cells from malignant primary tumors (see Figures **2d** and **3b**) can have clusters with tilted gradient lines **6**. In addition, if metastasis has occurred, the primary tumor clone can be identified by its tilted gradient line and its NPE in the metastatic site (see Figures **2d**, **2e**, **3b**, **3c**).

Furthermore, neoplastic cells can be characterized by having abnormal ratios of cells in the G_0/G_1 cluster **2** and G_2+M cluster **4**. For example, G_0/G_1 cells **2** clearly predominate over the G_2+M cells **4** in the normal human lymph node cells shown in Figure **2b**. In the ovarian cancer cells in Figure **4e**, however, there are nearly equal numbers of G_0/G_1 diploid cells **2** and G_2+M diploid cells **4**, indicating that the ratio of cells in the G_2 and M stage of the cell division cycle has become abnormal. Thus NPEs can be useful for identifying a neoplastic state in a perspective view.

These methods are applicable to any type of tissue subject to cancer, including lymph node (Fig. 2e), breast (Figs. 2c, 2d), colon (Figs. 3b and 3c), gastric (Fig. 4a), prostate (Fig. 4c), ovarian (Fig. 4d), lung (Fig. 4f), leukemia (Fig. 5b), as well as cervical and testicular tissue, pancreas, liver, brain and small intestine. Other tissues subject to these methods include brain, ovary, testes, bone and exfoliated circulatory tissue.

10 Devices for performing the methods disclosed above are also provided by the present invention. A device for determining the NPE of a cell can have means for measuring one or more BCs and measuring SDN, and means for determining the NPE. A block diagram of such
15 an embodiment of the device is shown in Figure 7. Specific measuring means are discussed in detail above, in the examples below, and in the publications and patents cited herein.

Each of the publications and patents cited
20 herein are hereby incorporated by reference. The term "comprising" as used herein, including its use in the body of the claims, is intended to be open-ended, thereby encompassing the recited elements or steps, as well as encompassing embodiments having additional elements or
25 steps. The following examples are intended to illustrate but not limit the invention.

EXAMPLE I: PREPARATION OF STAINED NUCLEI

The following examples illustrate the preparation of isolated nuclei from tissue samples.

A. From Solid Tissue

Isolated nuclei from solid tissue specimens were prepared as follows. Approximately 1 to 2 cubic millimeters of tissue was placed in a petri dish with
5 2 ml of the following nuclear isolation solution to isolate and stain the nuclei: 10 mg/ml 4',6-diamidino-2-phenylindole (DAPI) DNA stain (Sigma Chemical Co.; St. Louis, MO); 0.6% NP-40 (v/v) (Accurate Scientific & Chemical Co.; Hicksville NY); in isotonic phosphate
10 buffered saline solution at pH 7.2. The isolated stained nuclei were filtered through a 35 micron polypropylene filter (RATCOM, Inc.; Miami FL) and placed on ice. Using this method, samples of isolated, stained nuclei were prepared from a variety of solid tissue sources,
15 including lymph node, intestine, breast, colon, thyroid, ovary, prostate, stomach and surface epithelial cells from mouth.

B. Human Lymphocytes

Human lymphocytes were isolated from venous
20 whole blood using ficoll hypate, washed with isotonic saline, and adjusted to a concentration of about 1×10^7 lymphocytes/ml. Then, 0.1 ml of this lymphocyte solution was added to 1 ml of the nuclear isolation solution described above, resulting in a solution of lymphocyte
25 nuclei at a concentration of approximately 1×10^6 nuclei/ml. The stained nuclei were filtered through a 35 micron polypropylene filter and placed on ice.

C. Trout Red Blood Cell Nuclei (TRBC)

Nuclei from trout red blood cells were used as an internal standard. Trout (*Salmo gairdnerii irideus*) red blood cells (U.S. Fish Hatchery; Erwin TN) were
5 prepared into stock solutions as follows: 150 μ l of a commercially available TRBC solution (RATCOM Inc.; Miami FL) was added to 2 ml nuclear isolation solution described above, resulting in a stock solution of TRBC nuclei at a concentration of about 2×10^6 nuclei/ml.
10 The stained TRBC nuclei were then filtered through a 35 micron polypropylene filter and placed on ice.

EXAMPLE II: DETERMINATION OF NPEs

A. DNAnalyzer Flow Cytometer

The DNAnalyzer™ flow cytometer (RATCOM Inc.;
15 Miami FL) simultaneously analyzed the DNA content by fluorescence and the volume of each particle (ENV) as it passed through the measuring aperture of the instrument. The volume was determined by the Coulter Electronic Cell Volume principle described in U.S. Patent No. 2,656,508
20 to Coulter, and according to the teaching in U.S. Patent No. 4,818,103 to Thomas and Eggleston.

The DNAnalyzer used a unique equilateral triangle flow cell with a cross-section of 70 microns per side and 70 microns in length. The inlet and outlet were
25 also equilateral triangles in cross-section, with dimensions starting at 1 cm per side, and decreasing in size over 1 cm to 70 microns per side. The instrument employed epi-illumination and collection optics (i.e. collection optics from the same side of the sample as

excitation optics) with a 410 nm dichroic mirror to excite the sample and collect the fluorescence emission.

The excitation source was a 100-watt stabilized mercury arc lamp with a 30-micron spot size in the most uniform region of the arc. A UG1 filter selected the optimal excitation wavelength for the DAPI of 365 nm from the emission of the mercury arc lamp. The micro-objective had a 1.25 N.A. corresponding to a collection angle of 120 degrees for collection of the fluorescence emission. The DNAAnalyzer then used Digital Pulse Processing (DPP) to determine the peak value of the fluorescence emission and nuclear volume from each nucleus.

B. Sample Analysis

Samples of stained nuclei prepared as described in Example I were analyzed as follows. Approximately 50 μ l of the TRBC stock suspension was added to each sample as an internal DNA standard. The sample was then analyzed on the DNAAnalyzer at a flow rate of 60 to 120 nuclei/second and at least 15,000 events were collected. The data were collected at 8 bits of resolution for both ENV and DNA fluorescence, and presented in an FCS 2.0 standard file format (The Institute of Electrical and Electronics Engineers, Inc; New York NY).

The data were analyzed using Modfit version 5.11 (Verity Software House Inc.; Topsham ME). The two-parameter graphs were prepared using WinMDI version 2.7 (freeware available at <http://facs.scripps.edu>). The contour and perspective plots were analyzed in log mode with the interval set at 85%, smooth at 6, and threshold

at 0.5. The display resolution was 256 x 265 for the contour and perspective displays.

Calibration was performed for volume using 4 micron calibration spheres and the known value for trout RBC nuclei of 5 pg of DNA per TRBC nucleus.

C. Determination of NPEs

Normal cells were prepared as described in Example I and measured for nuclear volume (cubic microns) and DNA content (picograms DNA). NPEs were determined in terms of DNA/ENV:

	tissue type	pg DNA	cu. micron	NPE
	lymphocyte	7.75	21.3	0.364
	lymph node	7.60	21.8	0.348
	breast	8.14	23.6	0.345
15	colon	7.90	22.2	0.356
	intestine	7.81	21.8	0.358
	thyroid	8.29	24.3	0.341

Alternative formulas for NPEs were also determined:

	tissue type	ENV/DNA	DNA ² /ENV	(DNA + ENV)/ENV
20	lymphocyte	2.75	2.82	1.364
	lymph node	2.87	2.65	1.348
	breast	2.90	2.81	1.345
	colon	2.81	2.81	1.356
	intestine	2.79	2.80	1.358
25	thyroid	2.93	2.83	1.341

D. Graphical methods for determining NPEs

Examples of NPE contours for normal human tissue are presented in the figures for surface epithelial tissue from mouth (Fig. 1a), intestine (Fig. 1b), thyroid (Fig. 1c), lymph node (Fig. 2a), colon (Fig. 3a) and lymphocytes (Fig. 5a). A perspective view of Figure 2a is provided as Figure 2b to emphasize the relative heights of the peaks.

In each of the figures, internal standard TRBC appears as cluster 1. The values for cubic microns and picograms of DNA per channel accompany each of the axes. A line 5 passes through the origin and the G_0/G_1 peak 2 and any diploid G_2+M peak 4 visible. An NPE (ENV/DNA) can then determined to be the slope of the line 5. Alternatively, the inverse slope (DNA/ENV) can be used as the NPE.

EXAMPLE III: USING NPEs TO CHARACTERIZE CELLS

This example illustrates the use of NPEs to characterize cells, for example by distinguishing among normal and neoplastic cells, and cells in different stages of differentiation and apoptosis.

A. Human lymph node cells

For reference, Figure 2a shows an NPE contour for cells from normal human lymph node. The G_0/G_1 cluster 2 and diploid G_2+M cluster 4 are relatively circular and their centers are aligned on NPE line 5.

In contrast to Figure 2a, Figure 2c shows an NPE contour for cells from benign breast tumor. The normal cells 2, 4 have an NPE of about 0.145 (pg DNA/cu. microns), shown as normal NPE line 5. As shown, the shape of the G_0/G_1 cluster 2 has stretched vertically by the presence of aneuploid cells 2c. At the upper edge of the 2c cluster, the NPE can be as low as 0.023 (pg DNA/cu. microns). This change in the shape of the cluster is indicative of a neoplastic state. It should be noted that the gradient line 6 is essentially vertical in this figure, expressible as a nearly vertical slope of 1041 (change in volume/change in DNA). Also, the TRBC internal standard 1 is also stretched somewhat as an artifact, due to adhesion of foreign particles to the TRBCs.

Figure 2d shows an NPE contour for cells from a malignant primary breast tumor. As shown, the G_0/G_1 cluster 2 and diploid G_2+M cluster 4 are both elongated by aneuploid cells 2c and 2d. Significantly, the gradient lines 6 are tilted clockwise from the vertical, expressible as a relatively less vertical slope of 9.4. Thus, a change in the slope of the gradient line of a cluster can be indicative of an abnormal condition such as a malignant neoplastic state.

Figure 2e shows an NPE contour for metastatic cells from the primary breast tumor in Figure 2d. The two aneuploid clusters 2c and 2d from Figure 2d are clearly recognizable by the slope of the clusters, gradient slope and NPE slopes in the metastatic aneuploid populations 2c and 2d in Figure 2e. Thus, a change in the width, breadth or shape of the peak can be a significant indication of an abnormal cell condition such

as metastasis. Moreover, Figure 2e shows that aneuploid population 2d can be clearly discerned from the diploid G₂+M population 4.

Furthermore, in Figure 2e, the second aneuploid population 2d accounts for 3.4% of the cells, compared to 0.9% of the cells in the diploid G₂+M population 4, showing a high ratio of second aneuploid cells compared to G₂+M cells. Previously, tetraploidy could only be confirmed when the G₂+M population was greater than 15%. Here, this ratio can be observed with G₂+M levels lower than previously known in the field and could previously be observed from a profile of DNA fluorescence alone (see Hankey et al., Cytometry 14:472-477 (1993)).

B. Human colon cells

Figure 3a shows an NPE contour for cells from normal human colon.

Figure 3b shows an NPE contour for cells from a primary tumor. Significantly, the G₀/G₁ cluster 2 has become elongated by aneuploid cells 2c, and has a tilted gradient line 6. An aneuploid NPE line 5c is shown passing through the cluster of aneuploid cells 2c

Figure 3c shows an NPE contour for cells taken from at the end point of resection when surgically removing a metastasizing tumor shown in Figure 3b. As shown, the aneuploid cells 2c are still present and have a tilted gradient line 6, indicating the resection sample contains metastatic cells.

Figures 4a to 4f show NPE contours for other cancerous tissue sources: gastric (Fig. 4a and perspective view Fig. 4b), prostate (Fig. 4c), ovarian (Fig. 4d and perspective view Fig. 4e) and lung (Fig. 4f). Notably, Figure 4f of lung cancer cells shows a cluster of aneuploid S cells 3c and a cluster of aneuploid G₂+M cells 4c (TRBC standard omitted for scaling reasons), as well as a distinct aneuploid NPE line 5c passing through 2c, 3c and 4c.

10 C. Human lymphocytes

Figure 5a shows an NPE contour for normal blood lymphocytes. The G₀/G₁ cluster 2 is very slightly elongated vertically.

Figure 5b shows an NPE contour for leukemic lymphocytes. As shown, the presence of aneuploid cells 2c has altered the shape of the G₀/G₁ cluster 2, resulting in a tilted gradient line 6 and an NPE 5c for the aneuploid population clearly discernable from the diploid NPE line 5. The NPE contour provides a clear indication that aneuploid cells 2c are present in the sample.

Figure 5c shows an NPE contour for a cell sample of normal blood containing activated lymphocytes. In addition to the G₀/G₁ cluster 2, an additional cluster of activated lymphocytes 2e is discernable, with a distinct vertical gradient line 6e. Thus an NPE contour is useful for detecting cells having a different state of differentiation.

D. Cell cycle

The progress of the cell cycle can be traced in Figure 6a, which shows a contour of datapoints from a sample of a mouse cell line containing cells at various stages of the cell cycle. Cells at G_0 phase are shown in cluster 2a. A slight increase in nuclear volume is reflected by the cells in G_1 phase in cluster 2b. Once the cells begin replicating their DNA during S phase 3, the values for DNA fluorescence begin to increase until they are double the values for cells in G_0 phase, where they remain in G_2 phase 4a. Corresponding aneuploid cells in S phase 3c and G_2 +M phase 4c are shown in Figure 4f.

Upon initiating nuclear division, the cells in M phase 4b show a slight increase in nuclear volume. Once nuclear division is complete, the amount of DNA and the nuclear volume are both halved, as shown by the return to G_0 phase 2a. Thus, the progress of cells through each stage of the cell cycle is reflected in the clusters identifiable in Figure 6a. The cell cycle can also be seen in Figures 2a and 2b. Remarkably, the clusters remain aligned on the NPE line 5 throughout, demonstrating that the characteristic NPE for the cell line is maintained, even while the nuclei and cells are dividing.

E. Apoptotic cells

WEHI cells from a WEHI-231 murine B lymphoma cell line (Lombardi Cancer Institute; Wash. D.C.) were obtained untreated or treated with an apoptotic agent. As shown in Figure 6b, the NPE contour for nonapoptotic cells contains cells in various stages of the cell cycle,

as discussed above (clusters **2a**, **2b**, **3**, **4a**, **4b**). The apoptotic cells are in a distinct cluster **2f**, with a different NPE line **5f**. The separation between the nonapoptotic and apoptotic cells is highlighted in perspective view Figure **6b** (the origin is in the distant upper right corner). It can be observed that the volume of the nucleus remains relatively constant whether apoptotic or not, but the measured DNA decreases during apoptosis, whether due to decrease in DNA content or in DNA staining. This suggests that the volume of the nucleus during apoptosis is not maintained by the DNA content, but by other components, such as the nuclear matrix.

Although the invention has been illustrated by the examples above, it should be understood that various modifications can be made without departing from the spirit of the invention. Accordingly, the invention is limited only by the following claims.